samples were collected by cardiac puncture in sixty animals (thirty-eight of the first colony and twenty-two of the second colony) and the presence of rabies virus neutralizing antibodies was determined by SFIMT (Simplified Fluorescent Inhibition Microtest). The antigenic characterization of the isolates was made using a panel of monoclonal antibodies, which was produced and provided by Centers for Disease Control and Prevention (Atlanta, U.S.A), as established by Pan- American Health Organization for characterization of rabies isolates in Americas. Five bats were positive to rabies by FAT and MIT in each colony, 12% in the first colony and 12.5% in the second. However, two bats dead of the second colony were unsuitable for rabies diagnosis by traditional techniques and their brains were submitted to RTPCR with positive results, totalizing seven positives bats indicating 23.3% of rabies virus positivity. All blood samples analyzed presented neutralizing antibodies titers and sixteen animals (40%) from the first colony and two (4.6%) of the second presented titers ≥0.5 UI/mL. There was a positive correlation between the incubation period in mice and the antibodies titers observed in the bats. The samples with the higher incubation period for MIT (29 days) were from bats that showed the highest neutralizing antibody titer. Some bats negative by MIT and FAT and apparently healthy, presented high antibodies titers. The antigenic characterization showed only one antigenic profile (positive just to MAb C12) observed in previous studies with samples isolated in the same species of bats in Brazil. Genetic characterization was performed by sequencing of a fragment of N protein region and the rabies genetic lineage identified in these study were segregated with isolates obtained from other *Histiotus velatus* samples isolated in other regions of Brazil. These results show the importance of these methodologies for the epidemiological surveillance of rabies virus in bats and the necessity of the monitoring of bat colonies in parks and environmental reserves frequented by humans and where living other wildlife species as preventive actions of rabies control.

PT.062
INVESTIGAÇÃO DE CASO DE RAIVA EM FELINO, MUNICÍPIO DE SÃO PAULO, 2011

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Em 1969, no Município de São Paulo ocorreram 989 casos de raiva animal e cinco casos de raiva humana. Entre 1969 e 1973 (fundação do Centro do Controle de Zoonoses- CCZ/SP), o número de casos de raiva humana aumentou 2,2 vezes, o número de animais vacinados cresceu cinco vezes e observou-se um decréscimo dos casos de raiva animal, chegando a 56% do total ocorrido em 1969. A partir de 1981 não ocorreram mais casos humanos e entre 1983 e 2010 não foram registrados casos autóctones em cães e gatos. O perfil epidemiológico da raiva vem mudando em todo o Brasil, com restrição da área de circulação da cepa canina do vírus. Nas regiões em que a raiva foi controlada nos animais domésticos, os casos de raiva em humanos diminuíram e os animais silvestres passaram a representar um novo desafio. Em São Paulo a variante canina não tem sido mais detectada. Atualmente as variantes circulantes são relacionadas a quirópteros, ocorrendo anualmente, em média, dois a quatro casos em morcegos não hematófagos. Em 01/12/2011 o CCZ/SP foi comunicado de diagnóstico positivo para raiva de um felino, com histórico de contato com quiróptero e morte sem sintomatologia. O animal foi a óbito no dia 3/10/2011 e encaminhado no dia 04/10/2011 para a Faculdade de Medicina Veterinária e Zootecnia/USP com suspeita de envenenamento. A liberação do resultado positivo ocorreu em 01/12/2011. O felino, uma fêmea, castrada, dez anos, tinha livre acesso à rua e histórico de vacinação anterior a 2010. No imóvel situado em área estritamente residencial no Distrito de Moema, vivem cinco cães e 23 felinos. A região é bastante arborizada, com árvores que podem oferecer abrigo e alimento para diferentes espécies de morcegos, nas proximidades de um parque arborizado, Parque do Ibirapuera (à 750m de distância). Frente à confirmação da variante Desmodus rotundus/Artibeus lituratus desenvolveram-se ações de bloqueio em área de 500m de raio, a partir do foco. Foram realizadas visitas domiciliares, levantamento de abrigos, avaliação e orientação para encaminhamento médico de moradores e freqüentadores da casa que tiveram contato com o animal doente, vacinação contra raiva e identificação de todos os animais da moradia, com observação por 180 dias a partir do óbito do animal positivo. Todos os imóveis da área de abrangência foram visitados, totalizando 1.277 imóveis trabalhados, 769 fechados e 140 recusas. Houve distribuição de material educativo, e foram vacinados contra raiva 102 cães e 16 gatos, com histórico de mais de seis meses de vacinação, no raio de cobertura de foco. Os animais contactantes foram acompanhados pelo CCZ, no período de observação, mantendo-se saudáveis. Recomenda-se o implemento de ações de vigilância: – laboratorial; – das agressões; – de rumores e casos suspeitos de animais com morte súbita ou histórico de contato com quirópteros ou outros animais silvestres e a revisão de estratégias do controle da raiva devido à mudança da situação epidemiológica da doença no município.

PT.063
ANTIGENIC AND GENETIC STABILITY OF RABIES VIRUS AFTER CONSECUTIVE PASSAGES IN MICE AND IN CELLS

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Despite the recognized stability of the rabies virus (RABV), antigenic and genetic differences among strains isolated from different species have been found. Different factors may be involved in generating heterogeneity in RABV, including duration of infection, virus load and host immune response. This work was carried out in order to examine the antigenic and genetic stability of RABVs isolated from different natural reservoirs and to help the understanding of viral pathogeny after consecutive passages in different systems. In this study were used tree RABV strains, one isolated from canine, one isolated from haematophagous bat and the standard rabies virus strain (Challenge Virus Standard – CVS). These strains were submitted to five consecutive passages in mice and in cells. The consecutive passages in mice were made by intracerebral route, for that, groups of six mice were submitted to five inoculations with each one of the three RABV strains. The inoculated mice were observed daily and the dates of death were recorded. The consecutive passages in cells were made in "Neuroblast albino mouse" (N2a) cell lineage, for that, the strains were inoculated in suspension cells and incubated for 72 hours, subsequently, cells were frozen and thawed three times. Both mice and cell passages were submitted to antigenic and genetic characterization. The antigenic characterization was determined by indirect immunofluorescence (IIF) with a panel of eight monoclonal antibodies (Mabs) raised to RABV antigens. For the genetic characterization the total RNA was extracted with Trizol and submitted to reverse transcription-polymerase chain reaction (RT-PCR) with primers targeting the N and the G genes, the amplicons obtained were subjected to nucleotide sequence analysis. The RABV sequences were analyzed using Bioedit package.